Emission of volatile organic compounds from petunia flowers is facilitated by an ABC transporter

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Plants synthesize a diversity of volatile molecules that are important for reproduction and defense, serve as practical products for humans, and influence atmospheric chemistry and climate. Despite progress in deciphering plant volatile biosynthesis, their release from the cell has been poorly understood. The default assumption has been that volatiles passively diffuse out of cells. By characterization of a Petunia hybrida adenosine triphosphate–binding cassette (ABC) transporter, PhABCG1, we demonstrate that passage of volatiles across the plasma membrane relies on active transport. PhABCG1 down-regulation by RNA interference results in decreased emission of volatiles, which accumulate to toxic levels in the plasma membrane. This study provides direct proof of a biologically mediated mechanism of volatile emission.

Plants can direct up to 10% of photosynthetically fixed carbon toward biosynthesis of volatile organic compounds (VOCs) (1), which are low-molecular-weight lipophilic molecules (~100 to 200 Da) with high vapor pressures at ambient temperature. Plant VOCs are chemically diverse and mainly represented by terpenoids, fatty acid derivatives, benzenoids, and phenylpropanoids. VOCs play essential roles in pollinator and seed-disperser attraction, above- and belowground defense against herbivores and pathogens, plant-plant signaling, and allelopathy (2, 3). They also provide protection against abiotic factors such as high light, temperature, and oxidative stress (4). Moreover, emitted plant VOCs contribute to the production of secondary organic aerosols that act as cloud condensation nuclei, thus affecting atmospheric chemistry and climate (5, 6). Global annual emission of plant benzenoids alone is estimated to be millions of tons, leading to the formation of up to 10 Tg y−1 of aerosols (6). Plant VOCs are also of value to the floriculture, flavor, cosmetic, and fragrance industries.

In flowers and roots, VOCs are primarily produced in epidermal cells, from which they are released into the atmosphere (7, 8) and rhizosphere (9), respectively. In vegetative organs, VOCs are synthesized in glandular trichomes (e.g., in tomato, basil, and peppermint) (10) and reach the atmosphere through mechanical disruption, or they are made in the leaf mesophyll (11) and exit through the stomata (12) or cuticle (13). Irrespective of the biosynthetic site and emission route, at the subcellular level, VOCs must traverse the cytosol and cross the plasma membrane, hydrophilic cell wall, and sometimes the cuticle to exit the cell. It has been an open question how VOCs cross these barriers in intact cells to reach the environment or intercellular air spaces connected to stomata.

The default assumption that VOCs simply diffuse out of cells (14–17) may be true for small VOCs such as isoprene. However, modeling has predicted that emission of most VOCs driven solely by diffusion would lead to toxic VOC accumulation in membranes as a result of preferential partitioning of these compounds into lipid bilayers (17). Such accumulation could cause reorganization of cellular membranes that could lead to leakage of organelar or cellular content (18). This raises the hypothesis that VOC emission requires active transport (17).

We used Petunia hybrida flowers, which emit benzenoid and phenylpropanoid volatiles in a temporally regulated manner, to investigate whether VOC emission relies on protein-mediated export. Because certain nonvolatile hydrophobic compounds, such as waxes and diterpenes, are exported across the plasma membrane by adenosine triphosphate–binding cassette (ABC) transporters (19, 20), we searched petunia petal RNA sequencing (RNA-seq) data sets (21) for ABC transporter transcript sequences. We found one highly expressed candidate that was up-regulated 103-fold in petunia flowers between day 1 (bud stage) and day 2 after opening (postanthesis), the developmental stages with the lowest and highest VOC emission, respectively (22) (fig. S1). This gene encodes Petunia hybrida ABC subfamily G member 1, PhABCG1, a predicted plasma-membrane transporter of unknown function that is expressed almost exclusively in petals of open flowers and regulated by the ODORANT1 transcription factor (23), which controls VOC biosynthesis in petunia flowers (24).

To determine whether PhABCG1 is involved in VOC emission, we generated transgenic petunia RNA interference (RNAi) lines under control of the petal-specific linalool synthase promoter (25) to decrease PhABCG1 expression (fig. S2). Three independent lines with PhABCG1 transcript levels reduced by 70 to 80% (Fig. 1A) exhibited 52 to 62% decreases in total VOC emission (Fig. 1B), with a concomitant 101 to 157% increase in total internal VOC pools (Fig. 1B). In PhABCG1-RNAi flowers, emission of each individual VOC was reduced to a different extent (fig. S3A), and their corresponding internal pools increased (fig. S3B). Amounts of alcohol glycosides, representing 13% of the total VOC internal pool, were ~2.5 times as high in transgenic as in wild-type flowers (fig. S3C).

To directly test for transport activity, we expressed His-StrepII-tagged PhABCG1 (fig. S4) in Nicotiana tabacum Bright Yellow 2 (BY-2) suspension cells. Of 50 screened transformants, the majority gave a positive signal close to the expected molecular mass (75.2 kDa), as detected with antibodies against the His tag (fig. S5). Plasma membranes prepared from microsomal fractions were enriched in both PhABCG1 and the plasma-membrane marker H+-dependent adenosine triphosphatase (fig. S6). As a half-size ABC transporter, PhABCG1 possesses a single membrane domain and a single nucleotide-binding domain and may function as a homo- or heterodimer (19). Because no other half-size ABCG genes were up-regulated in flowers as much as PhABCG1 (fig. S1), we hypothesized that PhABCG1 functions as a homodimer. When subjected to size-exclusion chromatography, PhABCG1 (fig. S7) elutes before the 158-kDa marker (fig. S8), consistent with the predicted homodimer size (2 × ~75.2 kDa) in a detergent micelle (~70 kDa). To test for PhABCG1 transport activity, we chose methylbenzoate and benzyl alcohol as substrates because they are major VOC constituents emitted by petunia flowers (22). On incubation of control and transgenic BY-2 lines expressing PhABCG1 with 14C-labeled substrates, we measured cell-associated radioactivity corresponding to the difference between passive diffusion into and active transport out of the cells. The observed decrease in cell-associated radioactivity in transgenic BY-2 cells relative to control cells demonstrates that PhABCG1 transports methylbenzoate and benzyl alcohol out of cells (Fig. 2, A and C). Two additional transgenic BY-2
We identified 16 additional ABCG transporter candidates in our RNA-seq data sets (fig. S1). All had lower transcript levels than PhABCG1 on day 2 postanthesis, and only two candidates (Ph4681 and Ph5139) exhibited up-regulation on day 2 versus day 1, (fig. S1, A and B). Whereas expression of Ph4681 was found to be 0.5% that of PhABCG1, expression of Ph5139 was 17% that of PhABCG1. However, transient RNAi down-regulation of Ph5139 had no effects on VOC emission or internal pools (fig. S9).

Phylogenetic analysis of all petunia flower ABCG transporters identified by RNA-seq and half-size Arabidopsis thaliana transporters with known function revealed PhABCG1 to have one closely related petunia protein, Ph19708 (65% amino acid identity), and no Arabidopsis ortholog (fig. S10). Ph19708 expression was 0.2% that of PhABCG1 (fig. S11), indicating that it has little, if any, contribution to VOC emission. Two additional PhABCG1 homologs (Ph3519 and Ph9795, sharing ~45% amino acid identity with PhABCG1) clustered with three Arabidopsis ABCG transporters known for transporting wax precursors (AtABCG11, AtABCG12, and AtABCG13) (27) and clustered separately from PhABCG1 and Ph19708 (fig. S10). To check whether PhABCG1 contributes to wax precursor transport, we analyzed wax in wild-type and PhABCG1-RNAi flowers (fig. S11). We found no statistically significant differences in wax composition, quantity, and thickness, indicating that PhABCG1 is not involved in export of wax constituents.

To test our model prediction that accumulated VOCs have detrimental effects on membrane integrity (17), we stained petunia petals of 2-day-old wild-type and PhABCG1-RNAi flowers with two compounds having different modes of action. The first, propidium iodide, diffuses into cells and stains cell nuclei, consistent with damage to the plasma membrane. The second, fluorescein diacetate, passes through intact cell membranes and remains colorless until the acetate moiety is removed by intracellular esterases. The deacetylated fluorescein is fluorescent and unable to pass through plasma membranes (28). The second, fluorescein diacetate, passes through intact cell membranes and remains colorless until the acetate moiety is removed by intracellular esterases. The deacetylated fluorescein is fluorescent and unable to pass through plasma membranes (28).
staining when fed with high concentrations (15 to 30 mM) of benzaldehyde (fig. S12). We also found less fluorescein staining in PhABCG1-RNAi petunia flowers. (A) Confocal microscopy images of 2-day-old WT and PhABCG1 transgenic flowers stained with propidium iodide (scale bars, 50 µm). (B) Confocal microscopy images of 2-day-old WT and PhABCG1 transgenic flowers stained with fluorescein diacetate (scale bars, 20 µm). (C) Quantification of fluorescein intensity of fluorescein in WT and PhABCG1 A1 lines stained with fluorescein diacetate. Values are means ± SE (n = 19). ***P < 0.001 (Student’s t test).

Here we show that VOC emission from petunia flowers depends not on passive diffusion but rather on protein-mediated transport across the plasma membrane. Transporters may regulate emission of VOCs in other plants and in organisms throughout other kingdoms of life, where VOCs play important roles in chemical communication.

REFERENCES AND NOTES


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SUPPLEMENTARY MATERIALS

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Materials and Methods

Figs. S1 to S15

References (30–50)

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Active transport of aromas
Volatile organic compounds (VOCs) serve as invisible lines of communication among host plants, pathogens, commensals, community groups, and, with flowers, their pollinators. Studying petunia flowers, Adebesin et al. show that VOCs do not passively diffuse out of the cells but are actively shuttled across the plasma membrane by an ABC (ATP-binding cassette) transporter (see the Perspective by Eberl and Gershenzon). Disabling the transporter results in damage to the cell's membranes by intracellular accumulation of VOCs. Science; this issue p. 1386; see also p. 1334

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