

Ubiquitin and the Deconstruction of Synapses

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The remodeling of synapses is a fundamental mechanism for information storage and processing in the brain (1, 2). Much of this remodeling occurs at the postsynaptic density (PSD), a specialized biochemical apparatus containing neurotransmitter receptors and associated scaffolding proteins that organizes signal transduction pathways at the postsynaptic membrane (3–5). Easily visible in the electron microscope, the PSD is a disk-like structure ~30 nm thick and a few hundred nanometers wide, positioned opposite presynaptic terminals (see the figure, below).

During synapse maturation and in response to synaptic activity, the PSD undergoes remarkable structural changes (2), including growth (6), complexification, and perforation (7, 8). Such structural plasticity provides a physical basis for enduring changes in neural circuits and is thought to be mediated by alterations in the molecular composition of the PSD (2, 5). Indeed, stabilization or removal of neurotransmitter receptors and signaling proteins from the PSD has been proposed to account for long-term changes in synaptic strength (2, 9–14). Nevertheless, with few exceptions, the specific (and likely numerous) molecular changes that occur in the PSD in response to synaptic activity remain unknown. More important, understanding the patterns of molecular changes in large sets of PSD proteins, which ultimately encode the history of activation at the synapse, represents a level of analysis not previously attempted. It has been our goal to obtain a more complete understanding of the molecular underpinnings of neural circuit modification through a multiprotein analysis of activity-dependent changes in the PSD.

To examine activity-dependent changes in the PSD, we used quantitative protein profiling approaches, together with a modified procedure for isolating PSDs from cultured cortical neurons (15, 16). This experimental system allowed for strong and reproducible manipulation of activity levels across a relatively homogeneous population of synapses

using pharmacological tools. We found that changes in PSD composition are bidirectional, saturable, reversible, and involve multiple classes of PSD proteins (16). Surprisingly, the time course and magnitude of bidirectional change in PSD composition were remarkably similar across all protein classes (see online fig. 2). The striking resemblance among the patterns of protein accumulation and loss from the PSD suggests that large sets of postsynaptic proteins exist as functional or physical ensembles, and it implies that control over PSD composition may be governed by the incorporation or removal of certain key “master organizing molecules” that preserve stoichiometric relationships between PSD proteins.

The long-lasting changes in the molecular content of synapses we observed could

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arise by two general mechanisms: the incorporation of new proteins or the selective removal of existing synaptic proteins. For much of the past two decades, the prevailing model for enduring changes in synapse function and structure has been stimulus-dependent gene expression and protein synthesis (17, 18). Indeed, substantial evidence indicates that transcriptional events are critical for long-term activity-dependent plasticity (17, 19). In addition, several studies support a role for local translation of dendritic mRNAs in orchestrating long-lasting forms of learning-related synaptic plasticity (18). On the other hand,

considerably less attention has been given to the contribution of protein turnover to long-term structural and functional changes at synapses. To examine the effect of activity on dynamic PSD turnover, we measured the half-life or turnover rate of synaptic proteins using metabolic labeling pulse-chase analysis. Remarkably, we found a robust ongoing turnover of total PSD protein in control neurons that was accelerated by neuronal activity and slowed in inactive cultures (16). Much to our surprise, the turnover rate of total PSD protein occurred on the order of only a few hours, suggesting that the entire complement of synaptic proteins in mature neural circuits are replaced multiple times a day!

What causes this robust and regulated turnover of synaptic proteins? Using further biochemical approaches, we demonstrated that ubiquitin conjugation and proteasome-mediated degradation are the primary mechanisms for activity-dependent remodeling of the PSD. Specifically, we found that activity regulates de novo ubiquitin conjugation and turnover of postsynaptic proteins generally, and ubiquitination of certain scaffolding molecules specifically (16). Moreover, activity-dependent changes in PSD composition were completely dependent on the degradation of ubiquitinated proteins by the proteasome. Our findings therefore indicate that activity-dependent protein turnover joins stimulus-dependent gene expression and protein synthesis (17, 18) in engineering durable changes in the cellular machinery that governs synaptic morphology and function.



Organizing signal transduction. Electron micrograph of an excitatory synapse showing the PSD (red arrow). [Adapted from figure 1 in (3)]

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The modular multiprotein nature of these durable changes may be due to removal or turnover of master organizing molecules. Consistent with this notion, we have shown that the postsynaptic scaffolds Shank, GKAP, and AKAP79/150 undergo selective activity-dependent ubiquitination (16). Shank and GKAP are multivalent adaptors that bind to each other and to numerous additional proteins in the PSD (5, 20). In addition, AKAP79/150 anchors PKA and PP2B to complexes containing AMPA receptors or NMDA receptors and PSD-95 family members (21). The ubiquitin-dependent removal of one or more of these scaffolds could provide a mechanism for selectively destabilizing numerous associated proteins in the PSD complex, thereby accounting for the clusters of proteins coregulated by activity.

The PSD is, in essence, a proteinaceous signal-processing machine, with scaffolds, receptors, and enzymes comprising the various gears (3). Our findings indicate that the molecular components of this machine accumulate or disperse in reproducible patterns influenced by activity level and ubiquitination, raising the possibility that the

signaling properties of the PSD machine are similarly plastic. To address this question, we examined the effect of alterations in activity on downstream signaling to CREB and ERK-MAPK: two prominent signal transduction cascades organized by proteins in the PSD and involved in synaptic plasticity (4, 19, 22). Our results indicated that NMDA receptors at active synapses elicit augmented activation of CREB, whereas their counterparts at inactive synapses are selectively coupled to ERK-MAPK (16). This reciprocal regulation of CREB and ERK-MAPK pathways provides clear evidence that activity-dependent reorganization of the postsynaptic apparatus regulates multiple facets of synaptic signaling.

Thus, far from being an immutable structure, the PSD contains hidden dimensions of interconnected protein networks within which reside the molecular traces of experience. By demonstrating that activity controls the global composition of the synapse through ubiquitin-dependent turnover, our research provides a new conceptual framework for understanding and ultimately predicting functional changes in neural circuits.

References

1. R. Yuste, T. Bonhoeffer, *Annu. Rev. Neurosci.* **24**, 1071 (2001).
2. C. Luscher, R. A. Nicoll, R. C. Malenka, D. Muller, *Nature Neurosci.* **3**, 545 (2000).
3. M. B. Kennedy, *Science* **290**, 750 (2000).
4. H. Husi *et al.*, *Nature Neurosci.* **3**, 661 (2000).
5. M. Sheng, M. J. Kim, *Science* **298**, 776 (2000).
6. V. N. Murthy, T. Schikorski, C. F. Stevens, Y. Zhu, *Neuron* **32**, 673 (2001).
7. Y. Geinisman, L. deToledo-Morrell, F. Morrell, *Brain Res.* **566**, 77 (1991).
8. N. Toni *et al.*, *Nature* **402**, 421 (1999).
9. T. Meyer, K. Shen, *Trends Cell Biol.* **10**, 238 (2000).
10. G. G. Turrigiano, S. B. Nelson, *Curr. Opin. Neurobiol.* **10**, 358 (2000).
11. A. Rao, A. M. Craig, *Neuron* **19**, 801 (1997).
12. R. J. O'Brien *et al.*, *Neuron* **21**, 1067 (1998).
13. R. Malinow, R. C. Malenka, *Annu. Rev. Neurosci.* **25**, 103 (2002).
14. T. C. Thiagarajan, E. S. Piedras-Renteria, R. W. Tsien, *Neuron* **36**, 1103 (2002).
15. K. O. Cho, C. A. Hunt, M. B. Kennedy, *Neuron* **9**, 929 (1992).
16. M. D. Ehlers, *Nature Neurosci.* **10**, 10 (2003).
17. A. E. West *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* **98**, 11024 (2001).
18. O. Steward, E. M. Schuman, *Annu. Rev. Neurosci.* **24**, 299 (2001).
19. J. P. Adams, J. D. Sweatt, *Annu. Rev. Pharmacol. Toxicol.* **42**, 135 (2002).
20. T. Boeckers *et al.*, *J. Neurochem.* **81**, 903 (2002).
21. J. J. Michel, J. D. Scott, *Annu. Rev. Pharmacol. Toxicol.* **42**, 235 (2002).
22. B. E. Lonz, D. D. Ginty, *Neuron* **35**, 605 (2002).

2003 Grand Prize Winner

Michael Ehlers grew up in Grand Island, Nebraska, and earned his bachelor's degree in chemistry from Caltech in 1991. He went on to the Johns Hopkins University School of Medicine, where he was awarded M.D. and Ph.D. degrees in 1998 and where he also conducted postdoctoral research. Dr. Ehlers is currently assistant professor of Neurobiology and director of the Neuroproteomics Laboratory at Duke University. His research focuses on the interface between neuronal cell biology and the plasticity of neural circuits, with emphasis on protein trafficking and turnover mechanisms in dendrites. He has won several other awards in neuroscience and is a Scholar of the Ruth K. Broad Foundation.



Karel Svoboda, for his essay "Imaging Experience-Dependent Synaptic Plasticity in the Adult Neocortex in Vivo." Dr. Svoboda grew up in the Czech Republic and Germany and received his bachelor's degree in physics from Cornell University in 1994. As a graduate student in biophysics at Harvard University, he measured the tiny steps and forces produced by individual kinesin molecules. After being awarded his Ph.D. in 1994, he pursued postdoctoral work at Bell Laboratories, where his interests shifted to synaptic and dendritic function and plasticity. Dr. Svoboda started his own laboratory at Cold Spring Harbor Laboratory in 1997. Work in his laboratory focuses on experience- and activity-dependent plasticity in the cortex, probed with imaging, physiological, and molecular tools.

Satchin Panda, for his essay "Shedding Light on Non-Image-Forming Photoperception in Mammals." Dr. Panda was born and raised in India, where he earned his bachelor's degree in plant biology from Orissa University of Agriculture and Technology. He joined the graduate program at the Scripps Research Institute, where he studied the circadian oscillator mechanism in plants in the laboratory of Dr. Steve Kay. Since receiving his Ph.D. in 2001, he has pursued postdoctoral research in Dr. John Hogenesch's lab at the Genomics Institute of Novartis Research Foundation, San Diego. Here he uses genetic and genomic approaches to gain an understanding of the light input pathway and of circadian regulation of behavior and physiology in mammals.

Rudolf Cardinal, for his essay "Succumbing to Instant Gratification Without the Nucleus Accumbens." Dr. Cardinal was born in Norwich, UK, and grew up in Folkestone, UK. He studied medical sciences and neuroscience at the University of Cambridge, where he received his bachelor's degree in 1996 and then pursued intercalated courses in clinical medicine and surgery with a Ph.D. in behavioral neuroscience, supervised by Prof. Barry Everitt. He was awarded his MB BChir Ph.D. in 2001. His Ph.D. thesis examined the neuropsychology of reinforcement processes, including the contribution of the anterior cingulate cortex to Pavlovian conditioning and the neuroanatomy of impulsive choice. After qualifying, he worked as a house physician and surgeon in East Anglian hospitals and is now a neuroscience lecturer at Cambridge.

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